

A Practical Synthesis of 5-Lipoxygenase Inhibitor MK-0633

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Practical, chromatography-free syntheses of 5-lipoxygenase inhibitor MK-0633 *p*-toluenesulfonate (1) are described. The first route used an asymmetric zincate addition to ethyl 2,2,2-trifluoropyruvate followed by 1,3,4-oxadiazole formation and reductive amination as key steps. An improved second route features an inexpensive diastereomeric salt resolution of vinyl hydroxy-acid 22 followed by a robust end-game featuring a through-process hydrazide acylation/1,3,4-oxadiazole ring closure/salt formation sequence to afford MK-0633 *p*-toluenesulfonate (1).

Introduction

Leukotriene metabolism plays a central role in inflammatory diseases such as asthma, chronic obstructive pulmonary disease (COPD), and atherosclerosis. In particular, the activation of the enzyme 5-lipoxygenase (5-LO) and its associated protein, 5-LO activating protein (FLAP), initiates a cascade that transforms arachidonic acid into inflammatory leukotrienes. As part of a drug discovery program in our laboratories, MK-0633 *p*-toluenesulfonate (1) was identified as a potent and selective inhibitor of 5-LO (Figure 1). Herein we wish to report two practical, chromatography-free syntheses

MK-0633 p-toluenesulfonate (1)

FIGURE 1. Structure of MK-0633 *p*-toluenesulfonate.

that are suitable for the preparation of multikilogram amounts of 5-lipoxygenase inhibitor MK-0633.

Results and Discussion

Two convergent synthetic approaches to MK-0633 are illustrated in Scheme 1. The first approach that we investigated

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SCHEME 1. Retrosynthetic Analysis

$$\begin{array}{c} \text{Me} \\ \text{HO} \\ \text{CF}_3 \\ \text{Me} \\ \text{OHC} \\ \text{SO}_3 \\ \text{Me} \\ \text{HO} \\ \text{CF}_3 \\ \text{F} \\ \text{F} \\ \end{array}$$

SCHEME 2

$$F_3C \longrightarrow OEt \xrightarrow{(S)-BINOL} OEt \xrightarrow{Et_2Zn, EtMgCl} Me \xrightarrow{HO} CF_3$$

$$6 \qquad 10 \qquad (S)-6$$

$$60-75\% \text{ ee} \qquad 88-94\% \text{ ee} \qquad 97.9-99.3\% \text{ ee} \\ 49\% \text{ yield over 3 steps}$$

$$9a \qquad 9b: R = (S)-BINOL 9d$$

$$9c: R = H$$

and demonstrated on kilogram scale involved an asymmetric BINOL-ethylzincate addition to trifluoropyruvate followed by a late-stage reductive amination between 2-amino-1,3,4-oxadiazole 3 and aldehyde 4 as key steps. A second generation and complementary approach to MK-0633 involved a late-stage cyclization of mixed thiosemicarbazide 5. Both approaches required the development of robust syntheses of chiral α -trifluoromethyl α -hydroxy-acid intermediate 6. We established the second route described in this report as a potential manufacturing route to MK-0633.

Synthesis of Aminooxadiazole Intermediate. The synthesis of 2-amino-1,3,4-oxadiazole **3** began with organometallic addition to ethyl 2,2,2-trifluoropyruvate to form chiral ethyl hydroxy-acid **6** (Scheme 2). Few methods have been reported for asymmetric organometallic additions to α -keto esters. Furthermore, the challenging asymmetric synthesis of

chiral trifluoromethylated alcohols has received attention recently. $^{5-7}$

To install the tertiary alcohol center found in MK-0633, we discovered and developed an asymmetric ethylzincate addition to ethyl 2,2,2-trifluoropyruvate **8** (Scheme 2). Addition of EtMgCl to Et_2Zn at -20 °C led to complete formation of the corresponding chloromagnesium triethylzincate as ascertained by HNMR analysis. Subsequent addition of $Et_3ZnMgCl$ to a slurry of (S)-(-)-1,1'-bi(2-naphthol) [(S)-BINOL] in 1,2-dichloroethane at -20 °C led to formation of a chiral (S)-BINOL-zincate complex. Ethyl 2,2,2-trifluoropyruvate **8** was then added to this reagent over 4-6 h in order to obtain > 95% conversion to the desired adduct (S)-hydroxy ester **6** in 60-75% ee. Slow addition

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SCHEME 3. Synthesis of 2-Amino-1,3,4-oxadiazole 3

SCHEME 4. Synthesis of Aldehyde 4

of pyruvate 8 was required due to competing capture of the highly reactive pyruvate as a stable hemiketal 9b (R =(S)-BINOL) as ascertained by ¹⁹F NMR spectroscopic analysis. 10 In comparison, addition of pyruvate 8 over 5-10 min afforded at best 50% conversion to the desired ethyl adduct **9a**. Furthermore, the charge stoichiometry of Et₂Zn was found to be critical since excess reagent led to reduction of pyruvate 8 to form racemic alcohol 9d. 11 Pyruvate hydrate 9c, alcohol 9d, and (S)-BINOL were removed in the basic aqueous workup. It was essential to remove (S)-BINOL from the crude reaction mixture prior to ester hydrolysis. Omitting removal of (S)-BINOL led to poor yields of hydroxy-acid 6. Ester 9a was hydrolyzed in situ by treating the crude product with aqueous KOH at reflux. The crude acid 6 was solvent-switched to toluene and the enantiomeric purity was upgraded to 88-94\% ee via formation of a diastereomeric salt with (S)-1-(2-naphthyl)ethylamine. Salt break and recrystallization of the free acid in toluene gave (S)-6 in 49% yield over 3 steps and 97.9-99.3% ee.

Amidation of hydroxy-acid (S)-6 with HATU¹² activation and coupling with thiosemicarbazide 11 in THF gave hydrazide 12 in 91% yield (Scheme 3). A reslurry in acetone partially removed residual 1-hydroxy-7-azabenzotriazole (HOAt) and N, N, N, N'-tetramethylurea coupling byproduct and provided 12 in 99% yield, suitable for the oxidant-mediated cyclization. Other peptide coupling reagents gave inferior results in terms of yield and purity: CDI (40-59%

yield), EDC·HCl/HOBt (68-72% yield), TBTU¹³ (46% yield), PyBOP¹⁴ (68% yield).

Oxidant-mediated cyclization of thiosemicarbazide 12 with I₂/KI/aqueous NaOH proceeded to afford aminooxadiazole 3 in 66% yield (Scheme 3). Prolonged reaction time led to decomposition of 3 under basic conditions. Other oxidants gave inferior results: Na₂WO₄·2H₂O/H₂O₂ (decomposition), Br₂ (55% yield), HIO₃ (34% yield). Recrystallization of (S)-aminooxadiazole 3 from MTBE removed colored impurities and gave enantiomerically pure 3 as ascertained by chiral SFC analysis.

Synthesis of Aldehyde Intermediate. The synthesis of the aldehyde component began with the von Pechmann reaction between resorcinol **13** and *β*-keto ester **14** in methanesulfonic acid to afford hydroxy-coumarin **15** (Scheme 4). ^{15,16} Reaction with triflic anhydride gave triflate **16** and reductive carbonylation ¹⁷ with Pd(OAc)₂/dppp/Et₃SiH/Et₃N gave aldehyde **4** contaminated with the corresponding acid **17** (\sim 5A% HPLC) and reduced coumarin **18** (\sim 10A% HPLC) as determined by LC-MS analysis. These impurities were readily removed by a reslurry of crude coumarin aldehyde **4** in acetone.

Reductive Amination End-Game. Although a plethora of reaction conditions for imine formation were evaluated, we found the reductive amination to be a challenging step (Scheme 5). ¹⁸ The difficulty observed in the formation of the imine 20 was attributed to the poor reactivity of 2-amino-1,3,4-oxadiazole 3 since aldehyde 4 readily formed imines with more nucleophilic amines such as benzylamine. Reaction conditions involving azeotropic removal of water with benzene as solvent and 10 mol % pyridinium *p*-toluenesulfonate as acid catalyst for 18 h gave up to 80% conversion and 70% yield on small scale. Extended reaction times led to increased conversion but lower yields of imine suggesting that 20 was thermally unstable. Imine formation with coumarin

⁽¹⁰⁾ Hemiketals of ethyl 2,2,2-trifluoropyruvate prepared by reaction with the metal alkoxides of MeOH, 2-propanol, phenol, and BINOL exhibited a singlet with a chemical shift between δ –85.4 and –85.8 ppm by ¹⁹F NMR spectroscopic analysis with benzotrifluoride as internal standard (δ –61.5 ppm).

⁽¹¹⁾ In contrast, the reagent obtained from lithium triethylzincate, formed by reaction of ethyllithium with diethylzinc, and (S)-BINOL gave the opposite sense of stereoselectivity (52% ee).

⁽¹²⁾ HATU: 1-[bis(dimethylamino)methylene]-1*H*-1,2,3-triazolo-[4,5-*b*]pyridinium hexafluorophosphate 3-oxide: (a) Carpino, L. A. *J. Am. Chem. Soc.* 1993, 115, 4397. (b) Carpino, L. A.; El-Faham, A.; Minor, C. A.; Albericio, F. *J. Chem. Soc., Chem. Commun.* 1994, 201. (c) Carpino, L. A.; Imazumi, H.; El-Faham, A.; Ferrer, F.; Zhang, C.; Lee, Y.; Foxman, F. M.; Henklein, P.; Hanay, C.; Mügge, C.; Wenschuh, H.; Klose, J.; Beyermann, M.; Bienert, M. *Angew. Chem., Int. Ed.* 2002, 41, 441.

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⁽¹⁸⁾ Å thorough investigation of milder imine formation conditions with various dehydrating agents including Ti(OEt)₄, Ti(OtPr)₄, TiCl₄, (MeO)₃CH, MgSO₄, and CuSO₄ in toluene, 1,2-dichloroethane, or THF led to poor conversions to **20** (30–50%). Transimination via the *N*-TMS aldimine or N-H imine derivatives also proved fruitless.

SCHEME 5. Reductive Amination and Isolation of MK-0633 p-Toluenesulfonate

SCHEME 6. Hydrazide Synthesis

$$F_{3}C \longrightarrow OEt \qquad \begin{array}{c} 1) \text{ vinyIMgCl} \\ 2) \text{ LiOH} \\ \hline 73\% \text{ yield} \end{array} \longrightarrow OH \qquad \begin{array}{c} (S)\text{-phenylethylamine} \\ \hline 44\% \text{ yield} \end{array} \longrightarrow OH \qquad \begin{array}{c} Me \\ \hline 44\% \text{ yield} \end{array} \longrightarrow OH \qquad \begin{array}{c} Salt\text{-break} \\ \hline 44\% \text{ yield} \end{array} \longrightarrow OH \qquad \begin{array}{c} S \text{-phenylethylamine} \\ \hline 44\% \text{ yield} \end{array} \longrightarrow OH \qquad \begin{array}{c} S \text{-phenylethylamine} \\ \hline 44\% \text{ yield} \end{array} \longrightarrow OH \qquad \begin{array}{c} S \text{-phenylethylamine} \\ \hline 44\% \text{ yield} \end{array} \longrightarrow OH \qquad \begin{array}{c} S \text{-phenylethylamine} \\ \hline 44\% \text{ yield} \end{array} \longrightarrow OH \qquad \begin{array}{c} S \text{-phenylethylamine} \\ \hline 85\% \text{ yield} \end{array} \longrightarrow OH \qquad \begin{array}{c} S \text{-phenylethylamine} \\ \hline 85\% \text{ yield} \end{array} \longrightarrow OH \qquad \begin{array}{c} S \text{-phenylethylamine} \\ \hline 85\% \text{ yield} \end{array} \longrightarrow OH \qquad \begin{array}{c} S \text{-phenylethylamine} \\ \hline 85\% \text{ yield} \end{array} \longrightarrow OH \qquad \begin{array}{c} S \text{-phenylethylamine} \\ \hline 85\% \text{ yield} \end{array} \longrightarrow OH \qquad \begin{array}{c} S \text{-phenylethylamine} \\ \hline 85\% \text{ yield} \end{array} \longrightarrow OH \qquad \begin{array}{c} S \text{-phenylethylamine} \\ \hline 85\% \text{ yield} \end{array} \longrightarrow OH \qquad \begin{array}{c} S \text{-phenylethylamine} \\ \hline 85\% \text{ yield} \end{array} \longrightarrow OH \qquad \begin{array}{c} S \text{-phenylethylamine} \\ \hline 85\% \text{ yield} \end{array} \longrightarrow OH \qquad \begin{array}{c} S \text{-phenylethylamine} \\ \hline 85\% \text{ yield} \end{array} \longrightarrow OH \qquad \begin{array}{c} S \text{-phenylethylamine} \\ \hline 85\% \text{ yield} \end{array} \longrightarrow OH \qquad \begin{array}{c} S \text{-phenylethylamine} \\ \hline 85\% \text{ yield} \end{array} \longrightarrow OH \qquad \begin{array}{c} S \text{-phenylethylamine} \\ \hline 85\% \text{ yield} \end{array} \longrightarrow OH \qquad \begin{array}{c} S \text{-phenylethylamine} \\ \hline 85\% \text{ yield} \end{array} \longrightarrow OH \qquad \begin{array}{c} S \text{-phenylethylamine} \\ \hline 85\% \text{ yield} \end{array} \longrightarrow OH \qquad \begin{array}{c} S \text{-phenylethylamine} \\ \hline 85\% \text{ yield} \end{array} \longrightarrow OH \qquad \begin{array}{c} S \text{-phenylethylamine} \\ \hline 85\% \text{ yield} \end{array} \longrightarrow OH \qquad \begin{array}{c} S \text{-phenylethylamine} \\ \hline 85\% \text{ yield} \end{array} \longrightarrow OH \qquad \begin{array}{c} S \text{-phenylethylamine} \\ \hline 85\% \text{ yield} \end{array} \longrightarrow OH \qquad \begin{array}{c} S \text{-phenylethylamine} \\ \hline 85\% \text{ yield} \end{array} \longrightarrow OH \qquad \begin{array}{c} S \text{-phenylethylamine} \\ \hline 85\% \text{ yield} \end{array} \longrightarrow OH \qquad \begin{array}{c} S \text{-phenylethylamine} \\ \hline 85\% \text{ yield} \end{array} \longrightarrow OH \qquad \begin{array}{c} S \text{-phenylethylamine} \\ \hline 85\% \text{ yield} \end{array} \longrightarrow OH \qquad \begin{array}{c} S \text{-phenylethylamine} \\ \hline 85\% \text{ yield} \end{array} \longrightarrow OH \qquad \begin{array}{c} S \text{-phenylethylamine} \\ \hline 99.5\% \text{ yield} \end{array} \longrightarrow OH \qquad \begin{array}{c} S \text{-phenylethylamine} \\ \hline 99.5\% \text{-phenylethylamine} \\ \hline S \text{-ph$$

aldehyde **4** proceeded slower on scale and was attributed to the poor solubility of 2-amino-1,3,4-oxadiazole (*S*)-**3**¹⁹ and afforded at best 65% conversion. Reduction with NaBH₄ in MeOH gave crude **2** in 64.5% assay yield. The stoichiometry of NaBH₄ relative to imine **20** was found to be critical since excess hydride (>30 mol %) led to reduction of the coumarin ring to afford **19** and **21** as byproducts (Scheme 5). Removal of benzene from the reaction stream was done via azeotropic removal with MeOH, followed by azeotropic removal of MeOH with THF for the final step. Complete elimination of benzene was achieved during the final salt formation step. The removal of benzene was carefully monitored by HPLC analysis throughout the process.

A robust purification step was required to remove substantial quantities of alcohol **19** (up to 26A% HPLC) formed in the reductive amination step. We identified MK-0633 *p*-toluene-sulfonate salt (1) as a stable, crystalline, and bioavailable API form for safety assessment and clinical evaluation. Its crystal-

lization allowed robust purification from crude free base **2** obtained with purities ranging from 60A% to 74A% HPLC. Thus, treatment of crude MK-0633 free base with *p*-toluene-sulfonic acid hydrate in THF/heptane gave crystalline tosylate salt (1) as a white powder in 82% yield, 99.69 wt %, 99.81A% HPLC, 0.19A% HPLC benzyl alcohol **19**, > 99.9% ee, < 3 ppm residual Pd, PhH: not detected, < 2 ppm).

Although this initial synthetic route provided kilogram amounts of MK-0633 for preclinical evaluation, a number of attributes made it impractical and difficult to scale-up further than a few kilograms: (1) a highly work intensive and inefficient synthesis of the chiral hydroxy-acid (S)-6; (2) the expensive and not readily available peptide coupling agent HATU was used in the preparation of thiosemicarbazide 12; (3) a sensitive cyclization to 2-amino-1,3,4-oxadiazole (S)-3; and (4) a difficult reductive amination penultimate step. To address these shortcomings, a second generation and complementary approach to MK-0633 involved a late-stage cyclization of mixed thiosemicarbazide 5 (Scheme 1).

Synthesis of Hydrazide Intermediate. Because of both economic and operational considerations the stoichiometric BI-NOL-zincate addition was deemed not viable for long-term

⁽¹⁹⁾ Solid-state NMR analysis of (S)-3 indicated a different crystal form vs material recrystallized from iPAc/heptane.

⁽²⁰⁾ The formation of byproduct 21 obtained from conjugate reduction was attributed to residual palladium in the coumarin aldehyde.

SCHEME 7. Coumarin Isothiocyanate Synthesis

SCHEME 8. Synthesis End-Game

manufacturing of MK-0633. We therefore evaluated a number of chemo- and biocatalytic approaches for the synthesis of ethyl hydroxy-acid intermediate 6 (see the Supporting Information for discussion). Ultimately, we found that a simple route involving diastereomeric salt resolution provided (S)-6 (Scheme 6). Addition of vinylmagnesium chloride to ethyl 2,2,2-trifluoropyruvate 8 followed by in situ hydrolysis of the ethyl ester gave racemic acid 22 in 73% yield. Addition of 100 mol % (S)-phenylethylamine to a solution of acid in toluene at 90 °C gave the desired crystalline salt 23 in 43–45% yield and 92–94% ee. Attempted resolution with a number of other chiral amines gave inferior results. Hydrogenation of the olefin with either Pd/C or Pt/C afforded ethyl hydroxy-acid (S)-6 in 88% yield after enantiomeric purity upgrade via recrystallization from toluene. In stark contrast, performing the diastereomeric resolution under a wide variety of conditions with ethyl hydroxy-acid 6 afforded 66% ee at best (with D-alaninol as chiral amine). Activation of the acid to its corresponding imidazolide with CDI and addition to 35% aqueous hydrazine at rt gave hydrazide 24 in 78–82% isolated yield after crystallization from toluene.

Synthesis of Isothiocyanate Intermediate. Coumarin triflate 16 was converted to the corresponding coumarin nitrile 25 by using Pd₂dba₃/dppf as catalyst and NaCN as cyanide source (Scheme 7). The desired coumarin nitrile 25 was isolated in 91% yield after simple precipitation from the reaction mixture. Alternatively, we found that the cyanation proceeded in 97% yield by using a catalyst prepared from

Pd(OAc)₂/Et₂Zn/dppf. Attempted cyanation of triflate **16** in the absence of a catalyst either failed or led to competing hydrolysis of the sulfonate group.²¹ Reduction of the nitrile with Raney cobalt in the presence of ammonia under 300 psi of H₂ at 90 °C gave complete conversion to the desired 7-aminomethyl-coumarin 26. A modified protocol with Raney cobalt in AcOH/H2O as solvent was later devised due to instability of the coumarin amine free base in NH₃/MeOH. Formation of isothiocyanate 7 was initially performed with carbon disulfide and p-toluenesulfonyl chloride in THF.²² Due to serious safety concerns toward handling hazardous CS₂ in the pilot plant we sought alternative conditions for the formation of 7. We initially found that use of thiophosgene in CH₃CN/Et₃N gave a mixture of desired 7 and the corresponding coumarin thiourea dimer 27 (30-70%). Use of inorganic base in CH₃CN/water gave improved profile and yield. Ultimately we found that powdered CaCO3 gave best results affording isothiocyanate 7 in 84% yield on kilogram scale after simple precipitation of product via addition of water.²³ Under these reaction conditions < 1% of the corresponding coumarin thiourea dimer 27 was formed.

⁽²¹⁾ The corresponding 7-bromo-coumarin afforded nitrile **25** upon reaction with CuCN but required elevated temperatures (150 °C, 60 h) to reach complete conversion.

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Through-Process End-Game. The end-game chemistry proceeded efficiently without isolation of intermediates (e.g., a through-process; Scheme 8). Thus, acylation of hydrazide **24** with isothiocyanate **7** in THF at rt and cyclization of thiosemicarbazide **5** with TsCl/pyridine²⁴ was followed by aqueous workup, carbon treatment, and solvent-switch to isopropyl acetate (iPAc) to afford crude MK-0633 free base **2**. Addition of **2** (\sim 74A% HPLC) to a solution of p-toluenesulfonic acid monohydrate (1.0 equiv) in iPAc at 60 °C gave crystalline MK-0633 p-toluenesulfonate salt (1) (91.4% yield, 99.82A% HPLC) as a white powder.

Conclusion

In conclusion, we have developed a practical, chromatography-free route to MK-0633 featuring a through-process hydrazide acylation/1,3,4-oxadiazole ring closure/salt formation sequence to afford multikilogram amounts of MK-0633 p-toluenesulfonate (1). It is notable that although major efforts were directed toward developing practical asymmetric routes to chiral α -hydroxy-acid intermediate $\mathbf{6}$, in the end a simple diastereomeric salt resolution emerged to provide $\mathbf{6}$ with improved productivity and ease of operation.

Experimental Section

(2S)-2-Hydroxy-2-(trifluoromethyl)butanohydrazide (24). Hydroxy-acid (S)-6 (3.18 kg, mol) was dissolved in THF (8 L). The solution was added over 1 h to a stirred slurry of CDI (3.6 kg, mol) in THF (8 L). Caution: Gas evolution, ensure good venting of CO₂! After stirring for 1 h, the resulting homogeneous solution was added over 1 h to a stirred biphasic solution of 35 wt % aqueous hydrazine (3.9 L) in THF (8 L). The reaction was stirred at rt for 18 h and then diluted with iPAc (28 L). A portion of aqueous hydrazine separated from the organic layer (\sim 1.7 kg) was drained off. The organic layer was then washed with citric acid (20% aqueous, 15 L then 3 L) and saturated aqueous NaHCO₃ (10 L). The layers were cut and solid NaCl was added to the combined aqueous layers until saturation. The aqueous layer was then back-extracted with iPAc (2 \times 13 L). The combined aqueous layers were extracted with iPAc (5 L). The organic layers were combined, line-filtered, and concentrated. The solution was solvent-switched to toluene at 25 °C and the resulting gelatin-like slurry was filtered and washed with toluene (8 L) to give **24** as a crystalline solid (2.78 kg, 97.7A%, 96.3 wt %, 99.5% ee, 78% yield): mp 98–99 °C, $[\alpha]_D^{20}$ – 26.7 (c 1.69, MeOH); ¹H NMR (DMSO- d_6) δ 9.25 (br s, 1H), 6.61 (br s, 1H), 4.49 (br s, 2H), 2.02 (m, 1H), 1.68 (m, 1H), 0.82 (t, 3H, J =7.0 Hz); ¹³C NMR (DMSO- d_6) δ 165.4, 124.8 (q, J = 285 Hz), 77.3 (q, J = 26 Hz), 24.8, 6.7; ¹⁹F NMR (DMSO- d_6) δ -78.6. Chiral GC analysis: column, Restek Rt-gammaDEX-sa, 30 m × 0.32 mm, $0.25 \mu \text{m}$ film (RTx-1701); injector, split $12 \times$, cup linear with Siltek; inj., 1.0 µL at 230 °C; det., FID at 250 °C; carrier gas, He = $3.4 \,\mathrm{mL/min}$, flow 24 psi, 180 °C isothermal; (S)-hydrazide $t_{\rm R} = 7.9$ min, (R)-hydrazide $t_{\rm R} = 9.3$ min; HRMS calcd for $C_5H_{10}F_3N_2O_2[M+H]$ 187.0694, found 187.0688.

4-(4-Fluorophenyl)-7-(isothiocyanatomethyl)-2*H*-chromen-**2-one** (7). A visually clean 100 L round-bottomed flask equipped with a mechanical stirrer, a dropping funnel and a N₂ inlet was charged with coumarin amine hydrochloride **26** (4.68 kg, 15.31 mol) and powdered CaCO₃ (2.30 kg, 22.97 mol, 150 mol %). The solids were

suspended in CH₃CN (23.4 L) and water (18.7 L) at rt to afford a white slurry that stirred well. Mixing caused an endotherm to 9.2 °C. A solution of thiophosgene (1.40 L, 18.37 mol, 120 mol %) in CH₃CN (1.40 L) was added dropwise over 45 min. Caution: proper venting is required due to gas evolution! The internal temperature rose from 9.2 to 24.5 °C during the addition of thiophosgene and gave a thick cream-colored slurry. The batch was stirred for 15 min and HPLC analysis showed >99% conversion to isothiocyanate 7. Water (23.4 L) was added dropwise over 30 min and the slurry was filtered. The cake was washed with water $(3 \times 18.7 \, \text{L})$ and dried on the frit for 3 h. The solids were dried under high vacuum/N2 sweep at 30 °C. Isothiocyanate 7 was obtained as an off-white solid: 4.0 kg, 84% yield; mp 127–128 °C; ¹H NMR (DMSO- d_6) δ 7.62 (m, 2H), 7.49 (d, 2H, J = 8.0 Hz), 7.42 (dd, 2H, J = 9.0, 8.5 Hz), 7.36 (d, 1H, J = 8.5 Hz), 6.47 (s, 1H), 5.10 (s, 2H); 13 C NMR (DMSO- d_6) δ 163.9, 161.9, 159.4, 153.6, 153.5, 139.3, 131.0, 130.8, 127.3, 123.2, 118.1, 116.0, 115.8, 115.5, 115.2; 19 F NMR (DMSO- d_6) δ -112.8; HRMS calcd for $C_{17}H_{11}FNO_2S$ [M + H] 312.0495, found 312.0489. HPLC analysis: column, Zorbax Extend C18 (4.6 \times 150 mm, 5 μ); eluent 0.1% aqueous NH₄OH/CH₃CN; 5-40% over 8 min, hold 2.5 min, to 90% over 5 min, hold 5.5 min, post-time 4.5 min; flow = 1 mL/min; detection = 210 nm; temp = 40 °C; inj., $10 \mu L$, coumarin amine $26 t_R = 11.4 \text{ min}$, coumarin thiourea dimer $27 t_R = 16.0 \text{ min}$, coumarin isothiocyanate 7 $t_R = 16.3$ min.

 $(1S)-N-\{[4-(4-Fluorophenyl)-2-oxo-2H-chromen-7-yl]methyl\}-$ 5-[1-hydroxy-1-(trifluoromethyl)propyl]-1,3,4-oxadiazol-2-amine (2). A visually clean 100 L round-bottomed flask equipped with a mechanical stirrer, a thermocouple, and a N₂ inlet was charged with solid isothiocyanate 7 (2.00 kg, 6.43 mol) and hydrazide 24 (1.28 kg, 6.62 mol). The solids were dissolved in THF (32 L) and the amber solution was let stir at rt overnight. Internal temperature was 20 °C. HPLC analysis indicated > 99% conversion to the desired mixed thiosemicarbazide 5 after ~15 h: mp 198.1-198.9 °C; ¹H NMR (400 MHz, DMSO- d_6) δ 10.31 (br s, 1H), 9.75 (br s, 1H), 7.99 (br s, 1H), 7.61 (dd, 2H, J = 5.6, 8.8 Hz), 7.37 (m, 4H), 7.23 (d, 1H, J = 8.8 Hz), 6.91 (s, 1H), 6.40 (s, 1H), $4.85 \,(\text{m}, 2\text{H}), 1.98 \,(\text{m}, 1\text{H}), 1.75 \,(\text{m}, 1\text{H}), 0.90 \,(\text{t}, 3\text{H}, J = 7.2 \,\text{Hz});$ ¹³C NMR (125 MHz, DMSO-*d*₆) δ 163.8, 161.9, 159.7, 153.8, 153.7, 144.5, 131.1, 131.0, 130.9, 126.4, 123.5, 123.1, 116.9, 115.9, 115.8, 114.8, 114.3, 77.9 (q, J = 26.1 Hz), 46.2, 25.5; ¹⁹F NMR (375 MHz, DMSO- d_6) δ –77.8, –113.1. HRMS calcd for $C_{22}H_{19}F_4N_3O_4S$ [M + H] 498.1111, found 498.1114. IR (cm⁻¹, KBr pellet): 3303, 2980, 1701, 1617, 1543, 1510, 1420, 1374, 1282, 1239, 1194, 1005, 841; $[\alpha]_{D}^{20}$ –4.8 (*c* 1.08, EtOH). HPLC analysis: 4.6 mm ×150 mm Eclipse XDB Phenyl column, gradient elution (0.1% H₃PO₄/CH₃CN from 65:35 to 10:90 over 50 min, hold 10 min; run time = 60 min), flow rate = 1.0 mL/ min, detection = 210 nm, T = 25 °C; sample preparation = an aliquot was withdrawn and diluted with CH₃CN/water (75/25). A 10 μ L sample was injected, thiosemicarbazide 5 $t_R = 22.68$ min. Chiral HPLC: Chiralpak AS-RH (4.6 mm × 150 mm); isocratic method, CH₃CN/water = 37:63, flow rate = 1.0 mL/ min, detection = 210 nm, T = 20 °C; sample preparation = an aliquot was withdrawn and diluted with CH₃CN/water (37/63), a 5 μ L sample was injected: $t_R = 21.7 \text{ min (major)}, 18.8 \text{ min}$ (minor). The reactor was equipped with a reflux condenser and p-toluenesulfonyl chloride (1.47 kg, 7.72 mol) and pyridine (1.09 L, 13.51 mol) were charged. The batch was heated to reflux (internal temperature = 67 °C) for ~18 h. HPLC analysis indicated > 95% conversion to MK-0633 free base 2. The heat was shut off and the batch was cooled to 35 °C. The batch was line-transferred into 1 N aqueous HCl (30 L) and then diluted with iPAc (30 L). The layers were cut and the top organic layer was washed with water (2 \times 30 L). The crude solution was combined with a batch of the same size to afford a total of \sim 6 kg of MK-0633 free base in iPAc/THF. The solution was treated with Darco KB-B (1.5 kg, 25 wt %) for 2 h at 18-19.7 °C and

⁽²⁴⁾ It is noteworthy that the corresponding semicarbazide analogue afforded only 20% assay yield of MK-0633 under the same conditions. We recently reported on the dramatically improved reactivity of thiosemicarbazides vs semicarbazides in the synthesis of aminooxadiazoles: Dolman, S. J.; Gosselin, F.; O'Shea, P. D.; Davies, I. W. J. Org. Chem. 2006, 71, 9548.

JOC Article

then filtered on solkafloc. The filter cake was washed with iPAc $(3 \times 10 \text{ L})$ and concentrated. The batch was solvent-switched to $iPAc (3 \times 20 L): mp 92.9 - 94.3 \, ^{\circ}C; [\alpha]_{D} - 4.5 (c 1.08, EtOH); ^{1}H$ NMR (400 MHz, DMSO- d_6) δ 8.54 (t, 1H, J = 6.0 Hz), 7.60 (m, 2H), 7.38 (m, 4H), 7.31 (d, 2H, J = 8.4 Hz), 7.23 (s, 1H), 6.41 (s, 1H), 4.51 (d, 2H, J = 6.0 Hz), 2.08 (m, 1H), 1.97 (m, 1H), 0.88 (t, 3H, J = 7.2 Hz); ¹³C NMR (125 MHz, DMSO- d_6) δ 164.1, 159.6, 156.1, 153.7, 153.6, 143.8, 131.0, 130.9, 126.8, 123.5, 117.4, 116.0, 115.8, 115.4, 114.7, 73.7 (q, J = 26.1), 45.4, 26.1; ¹⁹F NMR (375 MHz, DMSO- d_6) δ -79.8, -113.1; IR (cm⁻ KBr pellet) 3309, 3068, 2945, 1701, 1617, 1510, 1420, 1372, 1329, 1280, 1238, 1190, 1160, 1108, 1079, 1014, 841. HRMS calcd for C₂₂H₁₇F₄N₃O₄ [M + H] 464.1233, found 464.1228. HPLC analysis: 4.6 mm × 150 mm Eclipse XDB Phenyl column, gradient elution (0.1% H₃PO₄/CH₃CN from 65:35 to 10:90 over $50 \, \text{min}$, hold $10 \, \text{min}$; run time = $60 \, \text{min}$), flow rate = $1.0 \, \text{mL/min}$, detection = 210 nm, $T = 25 \,^{\circ}\text{C}$; sample preparation = an aliquot was withdrawn and diluted with CH₃CN/water (75/25). A 10 μ L sample was injected. HPLC $t_R = 17.65$ min. Chiral HPLC retention time: major enantiomer $t_R = 14.00$ min, minor enantiomer $t_{\rm R} = 19.75$ min.

5-[1-hydroxy-1-(trifluoromethyl)propyl]-1,3,4-oxadiazol-2-aminium 4-Methylbenzenesulfonate (MK-0633 Tosylate, 1). The crude solution of MK-0633 free base 2 (5.4 kg according to HPLC after concentration) in iPAc (15.5 L) was diluted in iPAc(40 L) and was stirred at rt for 16 h. Stirring was stopped and solids settled rapidly at the bottom of the flask. The supernatant was filtered and analyzed by HPLC and ¹H NMR spectroscopy. HPLC assay of the solution indicated 54.4 kg at 9.72% wt in iPAc for a total mass 5.3 kg of free base 2. ¹H NMR analysis showed a 1:8.74 wt ratio of free base: iPAc. Thus the free base solution already contained 52.89 L of iPAc. p-Toluenesulfonic acid·H₂O (2.17 kg, 11.42 mol) was dissolved in iPAc (13.2 L). The slurry was heated to 60 °C and stirred until complete dissolution of pTsOH \cdot H₂O (\sim 20 min). The solution of free base 2 was added over 2 h to the hot solution of pTsOH·H₂O. After 20% of the crude MK-0633 free base solution was added the addition was stopped. MK-0633 tosylate 1 seeds (1 g) were added and the mixture was stirred for 10 min. Addition of crude free base was then continued. After the end of the addition, the batch was gradually cooled to rt over 2 h, then let stir at rt for 16 h and the slurry was filtered. The filter cake was suspended in iPAc (40 L), filtered, resuspended in iPAc (40 L) and filtered. The batch was dried under low vacuum on the frit for 4 h. The solids were then transferred in trays to a vacuum oven at 30 °C. The solids were dried to a constant weight under low vacuum/N₂ sweep. MK-0633 tosylate salt (1) was obtained as a white solid (6.64 kg, 91.4% yield): mp 164-165 °C; $[\alpha]_{D}^{20} - 0.86$ (c 10.0, EtOH); ¹H NMR (500 MHz, DMSO- d_6) δ 8.58 (1 H, t, J = 6.2 Hz), 7.62 (2 H, dd, J = 8.3, 5.4 Hz), 7.49 (2 H, d, J = 7.8 Hz), 7.47–7.38 (4 H, m), 7.33 (1 H, d, J = 8.3 Hz), 7.13 (2 H, d, J = 7.7 Hz), 6.44 (1 H, s), 4.53 (2 H, d, J = 5.6 Hz), 2.30 (3 H, s), 2.17-2.05 (1 H, m),2.03-1.93 (1 H, m), 0.90 (3 H, t, J = 7.37 Hz); ¹³C NMR (125) MHz, DMSO- d_6) δ 164.1, 162.9 (d, J = 246.8 Hz), 159.6, 156.1, 153.7, 153.6, 145.5, 143.7, 137.7, 131.1 (d, J = 3.5 Hz), 130.9 (d, J = 8.7 Hz), 128.1, 126.8, 125.4, 124.5 (q, J = 286.6 Hz), 123.5, 117.4, 115.9 (d, J = 22.0 Hz), 115.4, 114.7, 73.7 (q, J = 28.6 Hz), 45.4, 26.1, 20.8, 7.0; ¹⁹F NMR (375 MHz, DMSO- d_6) δ -79.7, -113.1; HRMS calcd for $C_{22}H_{18}F_4N_3O_4[M+H]$ 464.1228, found 464.1246. IR (cm⁻¹, NaCl thin film) 3324, 3010, 2977, 1735, 1716, 1618, 1510, 1428, 1215, 1178. HPLC analysis: eclipse XDB-phenyl column 4.6 mm \times 15 cm (0.1% aq H_3PO_4/CH_3CN 65:35 to 10:90 over 50 min, 1.0 mL/min, 210 nm, 25 °C); MK-0633 (1) $t_R = 16.86$ min. Chiral HPLC analysis: Chiralpak AD-H column 4.6 mm × 25 cm (EtOH/hexane 60:40, hold 15 min, 0.5 mL/min, 300 nm, 30 °C); (S)-enantiomer $t_R = 9.5 \text{ min}$; (R)-enantiomer $t_R = 11.5 \text{ min}$.

Supporting Information Available: Experimental procedures and characterization for compounds (*S*)-3, 4, (*S*)-6, 12, 15, 16, (*S*)-22, 25, and 26 and copies of ¹H, ¹³C and ¹⁹F NMR spectra of selected compounds. This material is available free of charge via the Internet at http://pubs.acs.org.